Force Generation and Cytoskeletal Structure of Single Platelets Molly Y. Mollica^{1,2}, Kevin M. Beussman³, Adithan Kandasamy³, Junmei Chen², Krithika Manohar³, José López^{1,2}, Wendy E. Thomas⁴, Nathan J. Sniadecki^{3,4}

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Figure 1. (A) F-actin of platelets on a fibrinogen (FBN)-coated coverslip show platelets with nodules or hollow structure while (B) platelets on von Willebrand Factor do not. (C) Frequencies of manually classified F-actin morphologies are significantly different (p < 0.0001, Pearson's Chi-squared test) and superresolution microscopy was used to observe these morphologies in higher resolution. (D) To reduce user bias and increase yield, the manual classifications were used to train a machine learning model, which predicts the F-actin morphology with 95.7% accuracy.



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Figure 2. (A) Black dots is a fluorescent pattern on a flexible polydimethylsiloxane (PDMS) surface. Without cells, the pattern is undisturbed. (B) With cells, pattern is displaced by cell forces. (C) The pattern is fabricated by microcontact printing fluorescent-bovine serum albumin. (D) Black dots is compatible with fixing and staining, making it possible to co-measure Factin morphology (left) and traction forces (blue arrows on right).

Conclusions

- Platelet **F-actin morphology is significantly** different on surfaces treated with fibrinogen versus VWF and these morphologies are identifiable via machine learning
- Platelets on VWF produce significantly more force, more force per area, and more centrally localized force Black dots enable co-measurement of F-actin structure and high-resolution single-platelet forces in a single image without restraining cell spreading. These observations would not be possible in low-resolution methods, methods that restrict cell spread size and shape, and/or methods that are incompatible with fixing and staining We hypothesize that these differences in F-actin morphology are GPIb-mediated; this hypothesis is under investigation
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